
5 STEPS TO A
GREAT
BLOOD
SMEAR

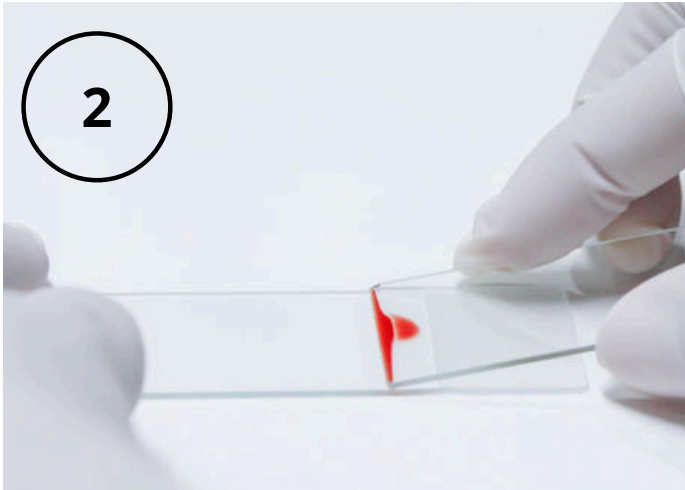
Prepare with 2 new, unused slides

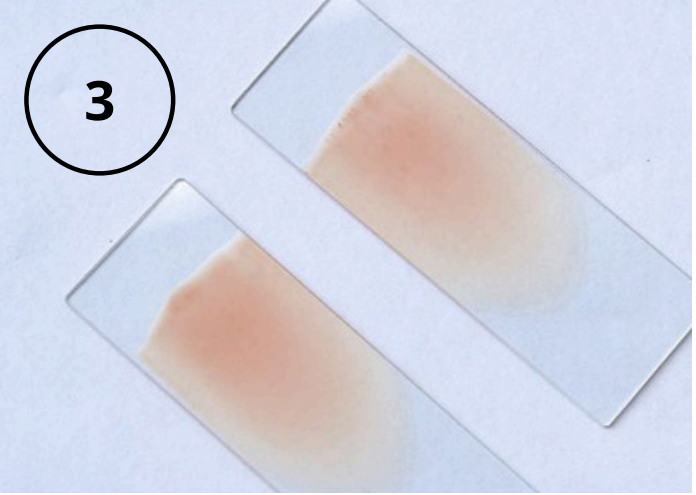
Place a very small drop of blood at one end of the slide

The ideal blood sample is directly and immediately from the patient as an anticoagulant can sometimes distort some cells.

Using your second slide as a slider, slide back until it hits the blood drop

Then move forward with a very smooth motion, continuing to the end of the slide





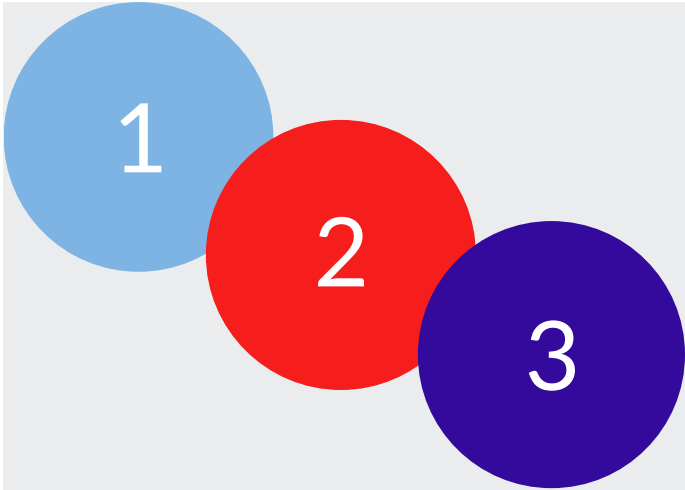
Your sample should end about 2/3 down the slides' length

This will create a '*feathered edge*' - this is where you will concentrate your analysis once stained

Allow your slide to air dry

Staining Order:

Stain 1 - Fixative
Stain 2 - Eosin Stain
Stain 3 - New Methylene Blue

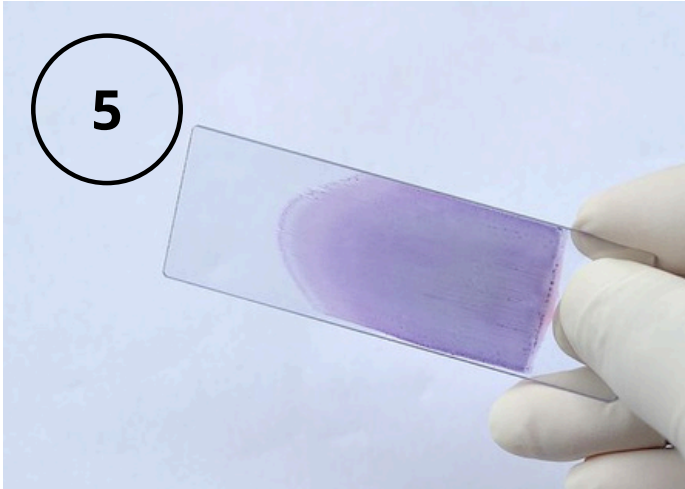


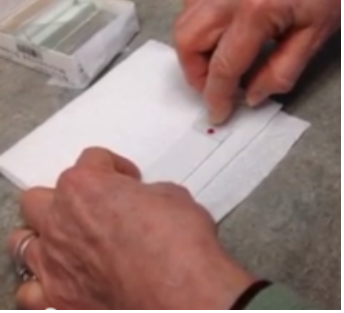
5 slow dips in each stain
(think counting 1-1000, 2-1000...)

Tap off excess stain on a paper towel between each stain

Rinse both sides of your slide with tap water

Allow to air dry (or use low-heat with a hair dryer)



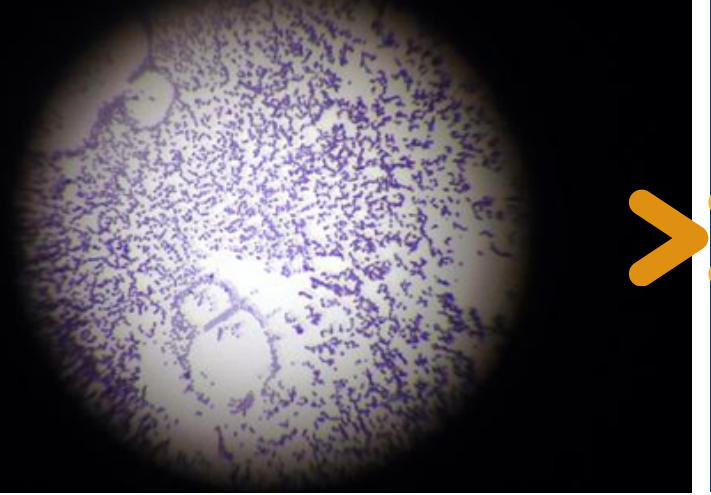


The Perfect Blood Smear? Practice Makes Perfect.

A swanky tool for making blood smears: Diamond Perfect Smear

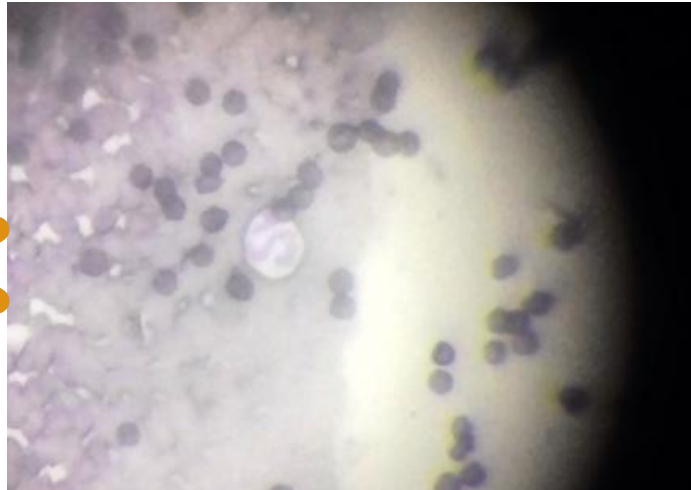
CELL COLLECTION

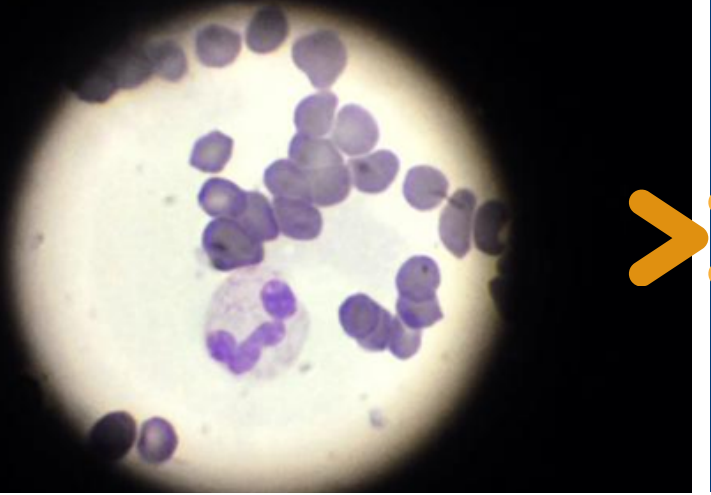
WHAT TO LOOK FOR ON YOUR SLIDES



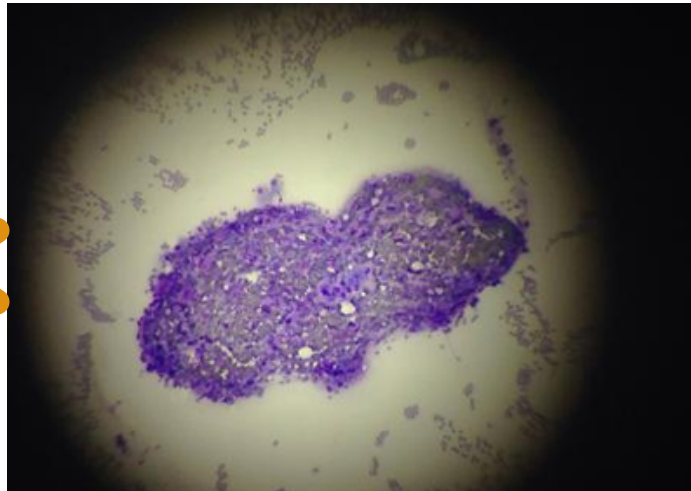
Slide on 10 x manification of a liver aspirate

Slide on 40 x magnification

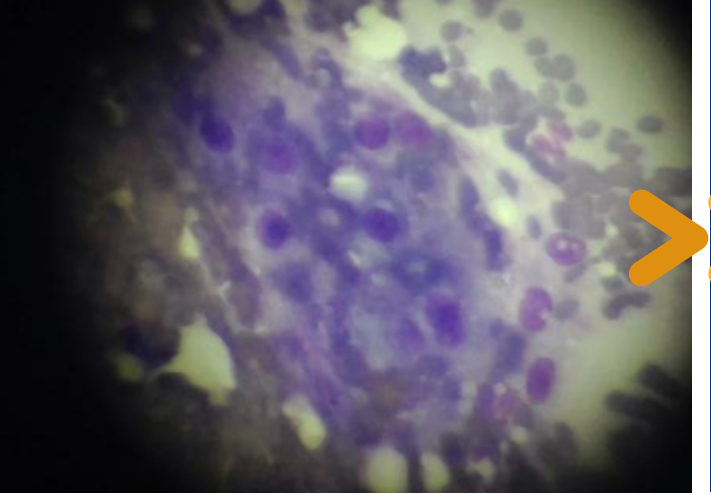




Slide on 100 x magnification with oil immersion

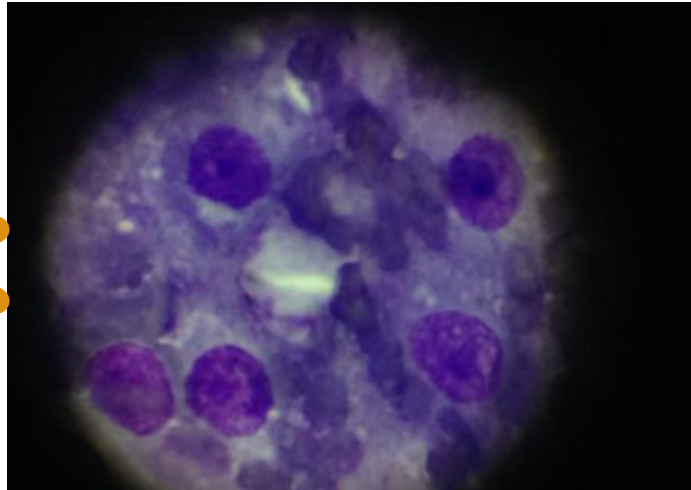


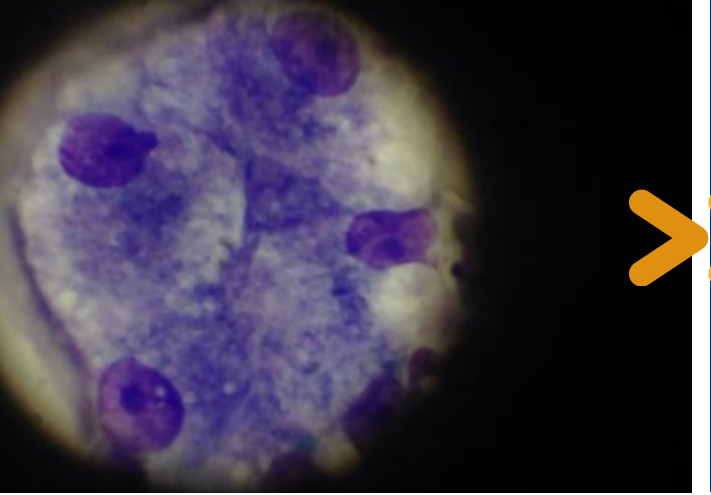
Hepatocyte cells at 10 x magnification.



Hepatocyte cells at 40 x magnification

Hepatocytes at 100 x magnification with oil immersion.





Another set of hepatocytes at 100 x magnification with oil immersion.

The key to good cytology stills and video, is to find nucleated cells in the slide first, then clumps of nucleated cells if possible and scan those regions.